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How to present and compare data obtained by particle tracking analysis and other related methods

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## Particle tracking analysis

Measuring sizes of individual particles

hydrodynamic diameter  $d_h > d_0$  due to diluent drag

Counting tracked particles in a given volume

investigated volume depends on size and refractive index

- Testing small volume of a sample
  - statistical process of estimating particle size distribution
  - volume tested about 100,000x smaller than sample volume



## **Concentration measurements**

• **Definition of**  $C_N$ : counts per volume [part/mL]

other defs: mass  $C_M$  [mg/mL] or volume  $C_V$  [ $\mu L/mL$ ]

- Typically  $\boldsymbol{C}_N$  is given for a range of sizes

e.g. between 100 nm and 1  $\mu$ m diameters

- Hence use of plots with size bins
  - bin widths not necessarily equal
  - some software do not support unequal bins (notably Excel)



## **Statistical considerations**

• Error in counting proportional to  $\sqrt{N}$ 

this applies to total counts and individual bin counts

Number of bins depends on expected errors

*N*=10,000 and 100 bins, average error 10 counts/bin or 10%

- To obtain "nice", smooth distributions:
  - decrease number of bins and use wider bins
  - use narrow bins to calculate parameters of a distribution



## **Example plots of size distributions**





# Binning

#### • Definition of a bin:

- $d \in [b_i, b_{i+1}]$  where i=1,...,N
- equivalent definition:  $b_i \le d < b_{i+1}$  (non-overlapping bins)
- typically  $b_1 = 0$  and  $b_N = max$  size to be measured
- Strange bins definition encountered:

		Average particles per mL CV,	, %	
29	particle diameter bin	Define the particle diameter bin range, for example: 100 nm $\leq d \leq$ 109 nm. The preferred bin width is 10 nm. The total range should cover from > 100 nm to < 2 µm. The range may be narrower, depending the range of the instrument being used. An example range is		
30	( 50 to 59) nm			
31	( 60 to 69) nm			
32	( 70 to 79) nm			
33	( 80 to 89) nm	given here, but may be modified as appropriate		
34	( 90 to 99) nm	for the measurement/instrument.		
35	( 100 to 109) nm		,	



## What is really plotted

- Concentration is just a rational number, also per bin
- Histogram uses area to represent a distribution
- With unequal bins natural measurable is:

density of particle size distribution (PSD)

units: number of particles per bin width and per investigated volume [part/nm/mL] total concentration ≡ area of a histogram



## **Dimensional analysis**

• When plotting concentration vs. size,

DO NOT connect points\*

- Total concentration is a sum of numbers, not an integral
- When plotting density of PSD, one can connect middle of bins // linear approximation
  - Total concentration  $\boldsymbol{C}_N$  is then

an area under the curve

\*There's no data between points – by definition of a bin



## **Practical procedure**

Create a list of measured sizes

typically diameters of nanoparticles are given in [nm]

Bin those sizes in specific binning

typically logarithmic binning

$$(b_{i+2} - b_{i+1}) / (b_{i+1} - b_i) = const$$

- Calculate density of PSD [part/nm/mL]; V<sub>0</sub> needed
- Plot as a histogram of density of PSD
- Calculate parameters of a distribution (AV, SD, D<sub>50</sub> ...)



## Example of real data in logarithmic binning



Concentration from 50 nm to 700 nm = area of histogram of density of PSD



## Real data vs. standards

• Most standards of size are mono-disperse

there are no standards for concentration...

Typically real life samples are poly-disperse

e.g. multiple sizes or continuous distributions

- Natural samples like sea water or blood:
  - highly poly-disperse with dominating small particles
  - Junge distribution  $N(d) \sim d^{-x}$  where  $x=3.5 \div 4$



## Fitted data problem (NanoSight FTLA)

- Fitting unknown distribution of sizes
- Assumed binomial(s) distribution
- Height (or area) of different peaks

is NOT conserved during fitting

(not invariant of any fitting procedure)

- Looks great but lacks numerical accuracy
- Artificial peaks can be created cf. van der Pol et al. J Thrombosis and Haemostasis, 12, 1182 (2014)



## Parametric description of distributions

- **Concentration**  $B_i = (b_{i+1} b_i)$   $N_{total} = \sum_{i=1}^{N} \frac{n_i}{B_i} B_i = \sum_{i=1}^{N} n_i$
- Average size  $d_{average} = \frac{1}{N_{total}} \sum_{i=1}^{N} n_i * (b_i + \frac{B_i}{2})$
- Standard deviation

$$SD = \sqrt{\frac{1}{N_{total}} \sum_{i=1}^{N} n_i * \left[ d_{average} - \left( b_i + \frac{B_i}{2} \right) \right]^2}$$

• **D**<sub>50</sub> 
$$D_k = \frac{1}{N_{total}} \sum_{i=1}^k n_i$$

definition 
$$D_k = 0.5 - k - b_k equal D_{50}$$



## D50 & mode depend on binning!





## Lies, damned lies, and statistics

• Parametric description using AV and SD

#### is not accurate or unique!

Anscombe's quartet – same AV and SD, various shapes

- Even higher moments do not give enough info
- Basic experimental question:

#### How similar are two measured distributions?



## **Kolmogorov-Smirnov statistics**



Comparing parameters:

 $d_{av} = 256 \text{ nm}, \text{ SD} = 145 \text{ nm}, \text{ CV} = 0.57$  $d_{av} = 163 \text{ nm}, \text{ SD} = 68 \text{ nm}, \text{ CV} = 0.42$ 

Non-parametric test: *Kolmogorov-Smirnov statistics* 

D <sub>A,B</sub>	alpha	<i>D</i> <sub><i>A,B,α</i></sub>	Reject?
0.2335	0.05	0.0338	yes



## **Anderson-Darling statistics**



Area between two cumulatives is a good measure of distance between two or more distributions with unknown shape.



## **Distance between distributions**

Normalize area between cumulatives

extreme case: a) particles at 10 nm, b) particles at 1000 nm

• **Distance is a number from the range** [0,1]

same distributions have distance 0

extreme case – distance 1



 If areas are calculated between distributions in same normalization, it can be a good measure



