



Elodie Ly-Morin<sup>1</sup>, Wilfrid Boireau<sup>2a</sup>, Patrick Ducouroy<sup>2b</sup>, Sophie Bellon<sup>1</sup>, Chiraz Frydman<sup>1</sup>

<sup>1</sup> HORIBA Scientific, Palaiseau, France,

<sup>2</sup> Clinical-Innovation / Proteomic Platform (CLIPP),

<sup>2a</sup> Institute FEMTO-ST, Université de Franche-Comté, CNRS, ENSMM, UTBM - F-25044 Besançon, France,

<sup>2b</sup> Centre Hospitalier Universitaire Dijon -CGFL – 1 rue du Pr Marion – 21000 Dijon, France

**Surface Plasmon Resonance (SPR) is an optical technique that offers label-free biomolecular analyses, providing information on kinetic processes (association and dissociation), binding affinity, analyte concentration and real time molecule detection. It has become a powerful tool for the analysis of biomolecular events involved in drug development, cancer research, and antibody screening, ...**

The phenomenon of SPR occurs when light interacts at the interface between a biochip and a liquid medium. It permits to follow modifications of the refractive index (or the reflectivity) in real time. Such modifications are induced by a biomolecular interaction between immobilized ligands (probe molecules) and captured analytes (target molecules). SPR monitors these changes of reflectivity to characterize the biomolecular events (such as binding and dissociation) occurring at the surface of the biochip in real time.

Surface Plasmon Resonance imaging (SPRi) technology offered by HORIBA Scientific takes SPR analysis a step further. The OpenPlex™ and XelPlex™ instruments (Figure 1) enable visualizing the whole biochip surface in real time using a video CCD camera.

This design allows biochips to be prepared in an array format; with each spot corresponding to a specific immobilized ligand. Up to several hundreds different molecules can be spotted using an automated spotter, opening the way to high throughput information for biomolecular interaction. The multiplexing capabilities of SPRi can meet with any experimental design without concession. Meanwhile, the sensitivity of SPRi is not compromised as analyte concentrations can be detected down to the nanomolar range.



Figure 1:  
HORIBA Scientific XelPlex™ system

The applications of SPRi are vast and include for example protein:protein<sup>1</sup>, D N A : D N A <sup>2,3</sup>, peptide:protein<sup>4</sup>, polysaccharides:proteins<sup>5</sup> or protein:cells<sup>6,7</sup> interactions.

The flexibility of the HORIBA Scientific instruments enables complex samples such as serum and plasma to be analyzed for clinical applications.

### General Overview of SPRi and Mass Spectrometry coupling

The coupling of SPRi biosensors and matrix-assisted laser desorption ionization mass spectrometry (MALDI-MS) is an innovative approach for biomarker discovery in biological fluids. It permits analytes captured by SPRi to be identified and characterized by their molecular weight and peptide sequence. SPRi-MS opens a new method of detection, quantification and structural characterization of proteins of interest. In the future, it could help better discriminate between sub-species within a family of biomarkers.

In this context, the complexity lies in the coupling of both techniques<sup>9</sup>. Most strategies require the elution of the bound analyte and its analysis by ESI- (electrospray ionisation) or MALDI-MS. This procedure has many drawbacks (analysis time, no multiplexing capabilities, decreased sensitivity, additional cross-contamination risks, etc...) which delayed the development of SPR-MS in the diagnostic field.

The open format of the HORIBA Scientific instruments makes MS coupling easier and faster. The possibility of direct MS analysis on the SPRi sensor was recently shown<sup>9</sup>. The SuPRA-MS platform (Surface Plasmon Resonance in arrays coupled with Mass Spectrometry) combines SPRi and MS in a single biochip. The biochip used for SPRi (SPRi-Slide) is directly transferred to the MS instrument. There is no need to neither elute nor re-deposit the bound analyte. The MS enzymatic digestion and the deposition of the MALDI matrix are performed directly on the SPRi-Slide. The latter is then directly placed on the MS plate holder (Figure 2).

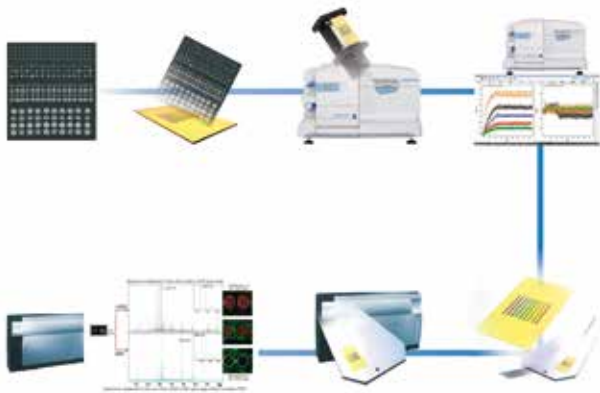


Figure 2: The SuPRa-MS platform

## Biomarker Capture and Identification using the SuPRa-MS Platform

A proof-of-concept study of SPRi-MS imaging coupling was performed for the detection of LAG3 recombinant protein in plasma. The solution fraction of this protein is a potential biomarker for breast cancer<sup>10</sup>. For this purpose, a mouse antibody (IgG2A) directed against LAG3 was immobilized on a SPRi-Slide using a dedicated surface chemistry compatible with MS analysis (NHS chemistry). Before injecting LAG3, rat serum albumin (RSA) was used to avoid non-specific binding on the surface of the biochip. Then, the specific interaction of LAG3 (added in plasma) and IgG2A was monitored using SPRi and images of the interaction were studied. Several femtomoles/mm<sup>2</sup> of LAG3 proteins were captured by SPRi. After direct processing on the biochip surface (enzymatic digestion and matrix deposition), the SPRi-Slide was analyzed using a MALDI-MS imager (Ultraflex, Bruker Daltonics). By showing the distribution of MS peaks specific of LAG3 and RSA respectively, it was possible to build the MS image of LAG3 spots (Figure 3) directly on the SPRi-Slide.

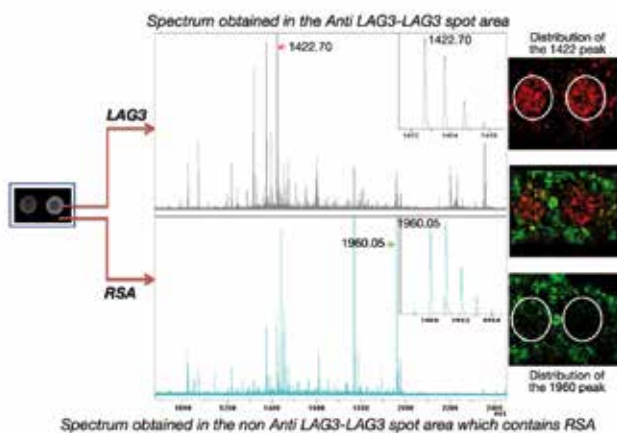


Figure 3: On-a-chip detection, identification and imaging LAG3 protein (potential marker of breast cancer) at 10 nM in human plasma through the SupRA-MS platform

The SuPRa-MS platform pioneers the combination of SPR imaging and MS imaging (MSi). It offers the possibility to gain spatially resolved information on the capture, sequence and molecular weight of clinical biomarkers.

## Conclusion

Multiplexed SPRi analysis using the HORIBA Scientific instruments provides rapid and high-throughput information in real time from up to several hundreds interactions in parallel. The technology is sensitive and does not require the use of labels. It can speed-up the workflow and reduce consumable costs during optimization processes. The coupling with MS analysis is straightforward and easier, which makes it a valuable tool for biomarker identification.

## References

1. Uzun and al. (2009) Production of surface plasmon resonance based assay kit for hepatitis diagnosis. *Biosensors and Bioelectronics*. 24(9): 2878-2884
2. Spadavecchia and al. (2009) New cysteamine based functionalization for biochip applications. *Sensors and Actuators B*. 143(1):139-143
3. Corne and al. (2008) SPR imaging for label-free multiplexed analyses of DNA N-glycosylase interactions with damaged DNA duplexes. *Analyst*. 133: 1036-1045
4. Prieto and al. (2009) Synaptonemal complex assembly and H3K4Me3 demethylation determine DIDO3 localization in meiosis. *Chromosoma*. 118: 617-632
5. Mercey and al. (2008) Polypyrrole oligosaccharide array and surface plasmon resonance imaging for the measurement of glycosaminoglycan binding interactions. *Anal. Chem*. 80(9): 3476-3482
6. Roupioz Y. and al. (2009). "Individual Blood-Cell Capture and 2D Organization on Microarrays" *Small* 2009, 5, No. 13, 1493-1497
7. Suraniti and al. (2007) Real-time detection of lymphocytes binding on an antibody chip using SPR imaging. *Lab Chip*. 7: 1206-1208
8. Boireau and al. (2009) Revisited BIA-MS combination: Entire "on-a-chip" processing leading to the proteins identification at low femtomole to sub-femtomole levels? *Biosensors and Bioelectronics* 24: 1121-1127
9. Bellon and al (2009) Hyphenation of Surface Plasmon Resonance Imaging to Matrix-Assisted Laser Desorption Ionization Mass Spectrometry by On-Chip Mass Spectrometry and Tandem Mass Spectrometry Analysis. *Anal. Chem*. 81: 7695-7702
10. Triebel and al (2006) A soluble lymphocyte activation gene-3 (sLAG-3) protein as a prognostic factor in human breast cancer expressing estrogen or progesterone receptors. *Cancer Letters*. 235(1):147-53.



**HORIBA**  
Scientific

[info.sci@horiba.com](mailto:info.sci@horiba.com)

[www.horiba.com/scientific](http://www.horiba.com/scientific)

**USA:** +1 732 494 8660  
**UK:** +44 (0)20 8204 8142  
**China:** +86 (0)21 6289 6060

**France:** +33 (0)1 69 74 72 00  
**Italy:** +39 2 5760 3050  
**Brazil:** +55 (0)11 5545 1500

**Germany:** +49 (0)89 4623 17-0  
**Japan:** +81 (0)3 6206 4721  
**Other:** +33 (0)1 69 74 72 00